



## Role of Banbara Nuts as Nutraceutical and its Potential in Green Synthesis of Nanoparticles: Ability of Bioactive Compounds

Joseph P Shaba <sup>1\*</sup>, Amusan TO <sup>2</sup>, Evans C Egwim <sup>3</sup>, Abdulkadir Abubakar <sup>4</sup>, Ossamulu I Famouse <sup>5</sup>

<sup>1</sup> Biology Unit, Air Force Institute of Technology, Kaduna, Nigeria

<sup>2,4</sup> Department of Chemistry, Air Force Institute of Technology, Kaduna, Nigeria

<sup>1,3,5</sup> Department of Biochemistry, Federal University of Technology Minna, Nigeria

\* Corresponding Author: **Joseph P Shaba**

### Article Info

**P-ISSN:** 3051-3405

**E-ISSN:** 3051-3413

**Volume:** 01

**Issue:** 02

**September – October 2025**

**Received:** 15-07-2025

**Accepted:** 17-08-2025

**Published:** 12-09-2025

**Page No:** 25-40

### Abstract

Bambara nut (*Vigna subterranea* (L.) Verdc.) is a vegetable crop that is easy to cultivate which has been underutilized over time. It is referred to as nutraceuticals due to its nutritional and medicinal values. The crop is tolerant to drought. Accordingly, it is important to explore the nutraceutical importance of this underutilized and neglected crop to safe guard and encourage its production for food as medicinal and nutritional source, particularly among the poor and rural societies in Africa and also to discuss its application and potentials in the synthesis of nanoparticles. This review specifically looked into the usefulness of bambara nut. The effect against microbes were also taken into consideration and studied to evaluate its nutraceutical advantages. Antimicrobial activities of bambara nut extracts have been observed in clinical settings against a variety of bacteria, *Candida albicans* and *Aspergillus niger* which was found to also possesses antioxidant properties. Bambara nut possess high bioactive compounds which include phenols and flavonoids that possess special chemical ability that can both effectively serve as capping and reducing agents on nanoparticles. Green synthesized zinc and silver oxide nanoparticles utilizing the post-harvest leaves of bambara nut and their antioxidant, anti-inflammatory and antimicrobial potentials have been reviewed. Likewise, the effect of nano-structured shell of bambara nut as filler have also been discussed. The potential applications of bambara nut as nutraceuticals and in green synthesis of nanoparticles are reviewed in this paper.

**DOI:** <https://doi.org/10.54660/IJABRN.2025.1.2.25-40>

**Keywords:** Bambara nuts, Nutraceuticals, Nanoparticles, Bioactive compounds.

### 1. Introduction

The world's legume plants are valued for their nutritional content and economic importance<sup>[1]</sup>. Despite being known as the meat for a poor man, legumes such as bambara nut (*Vigna subterranean*) are essential to human nutrition and health, particularly in impoverished remote sub-Saharan African communities<sup>[2]</sup>. This is due to their high protein and carbohydrate content as well as the various phenolic compounds they release when consumed, whether cooked or raw. Proteins and peptides found in legume seeds may be categorized as nutraceuticals<sup>[3]</sup>.

Because it has implications for the development of medications and dietary supplements for human wellness and health, the study of phytochemicals and mineral content in medicinal plants has gained interest in the pharmacological and nutritional science sectors<sup>[15]</sup>. Nutraceuticals are synthetic materials or chemical compounds (often found and concentrated in the form of capsule, pill, or powder) which can offer wellbeing or therapeutic advantages to human, and also promote the prevention of disease and their treatment<sup>[4]</sup>. Nutraceuticals are also defined by Sasi<sup>[5]</sup> as any functional food extract that has advantages for human health and medicine. Additionally, phytochemicals were described by Lopez-Gutierrez *et al.*<sup>[6]</sup> as substances found in plants which are non-nutritive and have the ability to prevent disease.

Recently, the majority of researchers' attention has focused on plant-based nutraceuticals since they include a wide range of phytochemicals, some of which are yet to be identified, that have been shown to have health advantages and the ability to prevent disease. Likewise, an eco-friendly procedures for nanoparticles' synthesis that is gaining popularity in the fields of biochemistry, biology, and chemistry technologies is called "green synthesis." This trend has a number of roots, one of which is the demand for more environmentally friendly ways to offset the increased expenses and energy demands of chemical and physical processes. This is why researchers look for less expensive synthesis techniques. The other reason is that standard techniques of synthesizing nanoparticles typically involve multiple procedures in the process of synthesis, which involves treatments with heat, and use harmful reductants like sodium borohydride or hydrazine, which frequently result in hazardous by-products. The goals of green chemistry are to reduce waste, use less energy, use renewable resources, and use risk-reducing techniques. The three primary ideas for creating nanoparticles using a green synthesis strategy include selecting a solvent medium (water is preferred) and using an environmentally friendly capping and reducing agents to stabilize the nanoparticles<sup>[7]</sup>. Then, it seems that the biological systems are the best factory for achieving these kinds of natural chemical conditions which can be found also in legumes such as bambara nuts. The use of plants to produce nanoparticles has sparked a lot of research in the field of green synthesis in more recent times<sup>[8]</sup>. Plant extracts contain bioactive compounds that can serve as capping and reducing agents to create stable, shaped nanoparticles. Enzymes, alkaloids, Proteins, phenolics, polysaccharides, amino acids, terpenoids, flavones, and alcoholic compounds are among the main substances that influence the capping and reduction of the nanoparticles. But reports have also included chlorophyll pigments, quinol, caffeine, linalool, methyl chavicol, eugenol, ascorbic acid, theophylline, and other vitamins<sup>[9–16]</sup>. Nontoxic Bioactive compounds, such as the phenols and flavonoids, possess a special chemical ability that can both effectively serve as reducing and capping agents on nanoparticles, preventing them from clumping together. Metals can bind to the carboxyl and hydroxylic groups present in the bioactive compounds<sup>[17]</sup>.

When compared to major income crops, bambara nuts are comparatively underutilized and are typically connected with small-scale subsistence farming, where women are the primary processors and producers<sup>[18, 19]</sup>. The lack of knowledge on how to improve seed systems, processing, agronomic techniques, and utilization—particularly with regard to the synthesis of nanoparticles using bambara nut are among the obstacles to the full potential of bambara nuts. Given the abundance of bioactive chemicals found in bambara nuts, this work explores their potential involvement in the eco-friendly synthesis of nanoparticles and the methods used while providing an outline of their nutraceutical usefulness.



**Fig 1: Bambara nut**

## 2. Description of Bambara nut and their cultivation

A natural crop that is extensively grown throughout most of sub-Saharan Africa is the bambara nut. The west, central, east, and southern subregions of the continent are among the agro-ecological zones where it is extensively farmed<sup>[20–23]</sup>. Bambara nut is an indigenous legume from Africa that is a member of the Faboideae and Fabaceae families, respectively. In the continent, bambara nuts have some common names such as Nzama in Malawi, Ntoyo in Ci Bemba, or Katoyo in Zambia, Njugo in South Africa. In Nigeria, it is known as Okpa (the Igbo speaking half) and Gurujia (the Hausa speaking section)<sup>[24]</sup>. According to Stephens<sup>[25]</sup>, the bambara nut is also referred to as the underground bean, baffin pea, voandzou nzama (in Malawi), and indhlubu. The annual leguminous plant known as the bambara nut has a robust taproot, trifoliate leaves, and a long, green petiole<sup>[26, 27]</sup>. The blooms are carried on long, hairy peduncles that emerge from the stem nodes<sup>[28]</sup>. The plant produces 3.0–3.7 cm-long, oval or spherical pods<sup>[9]</sup>. The landraces of Bambara nuts are made up of numerous genotypes with varying capacities to withstand environmental stress<sup>[27, 9, 30]</sup>. The plant takes 110–150 days to reach full maturity in well-drained soil, 5.0–6.5 pH sandy loams, with a mean temperature of 20–28°C<sup>[27]</sup>. The color of the seed can range from red, brown, or black and occasionally be speckled with many colors<sup>[22, 11]</sup>. Before they are eaten, the hulls are frequently removed. The removed shells are considered a waste material. Nonetheless, research by Klompong and Benjakul<sup>[18]</sup> has demonstrated that the hulls are a source of useful chemicals and nutraceuticals. Because of its resilience to severe environmental stress including low and variable rainfall and soils lacking in nitrogen, it is free from total crop failure<sup>[10, 12, 13]</sup>. The plant can thrive in humid regions and low-nutrient soil circumstances, where most other non-leguminous plants might not be able to survive<sup>[11]</sup>. According to Tibe *et al.*<sup>[14]</sup>, bambara nuts are a drought and disease-resistant crop that can help minimize pests in agricultural settings. Its capacity to fix atmospheric nitrogen through the process of biological nitrogen fixation reduces the need for costly, environmentally harmful synthetic fertilizers or manures, particularly in African's nutrient-poor soils, where resource-poor farmers cultivate the crop. Work

by Mohale *et al.* [15] on the symbiotic nitrogen nutrition effectiveness of the plant growing in the Lowveld region (Mpumalanga Province) of South Africa provided evidence for the nitrogen-fixing capacity of banbara nuts. The researchers discovered that the Banbara nut provided Nitrogen (33–98%) to the poor nutrient of the soils that are in the area and was found to be highly dependent on symbiotic fixation (4–200 kg N-fixed ha<sup>1</sup>) for its Nitrogen nutrition [15]. The use and cultivation of banbara nuts is of interest in sub-

Saharan Africa's arid savannah regions. 90% of farmers in Botswana (the bulk of them are women) grow banbara nuts. Of these, 63% grow the crop for personal use, 12% grow it for financial gain, and 25% grow it for both profit and food [16]. Given the crop's resilience to extreme environments, including drought and low-nutrient soils in the event of climate change, it is viewed as a crop capable of mitigating poverty and poor nutrition, particularly in impoverished sub-Saharan African's rural communities.



**Fig 2:** Cultivation of Banbara nuts

### 3. Nutritional importance of Banbara Nut

Farmed locally by the rural population, bambara nuts are source of protein for consumers and are primarily cultivated by them to feed their immediate families in most of the countries in sub-Saharan African region. According to study, the seeds of this crop may include 49–63.5% carbs, 15–25% protein, 4.5–7.4% fat, 3.2–4.4% fiber, 0.7% ash, and 0.01% cholesterol per 100 g dry weight [24, 11, 29]. Mwale *et al.* [18] noted that because the protein content is similar to that of other legumes, it is considered a valuable addition to diets based on cereal. The Banbara nut are cultivated below the ground, and the pods is hard and wrinkled when dried which contain at least one or two seeds. As a functional food, the dried seeds can be boiled and eaten. Additionally, the food crop can be eaten immature as a snack after roasting or steaming with a salt tincture [25] or matured as porridge [22]. The seeds are nutrient-dense and a complete food due to their chemical makeup [12, 31, 32]. The Banbara nut are produced below the ground, and the pods contain minimum of one or two seeds which are observed to be wrinkled and hard when dried. As a functional meal, the dried seeds can be cooked and eaten. Furthermore, the food crop can be eaten as porridge when it is mature [22] or as a snack when it is immature and has been roasted or sweltered with a salt tincture [25]. The seeds are a wholesome and complete diet because of their biomolecules [12, 31, 32]. Furthermore, the food crop can be consumed as porridge when matured [22] or the immature form as a snack after roasting or sweltering with a

tincture of salt [25]. The biomolecules of the seeds make them nutritious and a complete food [12, 31, 32]. Sesay [30] further stated that Banbara nuts are carbohydrates and protein sources to consumers and also provides extra cash for rural subsistence farmers in the majority of West African nations. Numerous researches have reported on the seed's biochemical composition [31–36]. Some regions of the Congo still harvest oil from the seed, despite the fact that it is said to contain little fat [37]. In East Africa, bambara nuts are roasted, pulverized, and added to cuisine without any sauces [37, 38]. Banbara nut flour is used to make part of the bread in Zambia [39]. The nuts are used in Botswana to make stiff porridge and pastries [34, 36]. Banbara nut flour is also used in Nigeria to make steamed foods like "okpa," [37]. Murevanhema and Jideani [40] defined "okpa" as a cooked substance, similar to gel, made from banbara nut paste, wrapped in banana leaves, and allowed to bubble until ready to use. Furthermore, the use of matured black landrace seed is used for therapeutic purposes [26]. The flavor and content of vegetable milk made from banbara nuts was found to be the best; also, people liked the lighter color of this milk over that of cowpea, soybean milk and pigeon pea [12]. Antioxidants are generally reported to be present in most legumes [40]. Antioxidants have the ability to preserve food quality by preventing the oxidative lipids degradation [41]. Additionally, Adelakun *et al.* [42] described antioxidants as components that shield human tissues and cells from free radicals (reactive nitrogen /oxygen species, or RNS/ROS). Antioxidants are abundant in most

leguminous nuts' seed coats and there have been reports of differences in antioxidant levels between the light- and dark-seeded landraces of banbara nuts [40]. Onyilagha *et al.* [43] stated that the antioxidants in the black and red seed coat of banbara nuts contain antioxidants whose mechanisms of antioxidation differ from those of other leguminous plant and were found to possess more nutrients and minerals than the light-seeded landraces. In Botswana, the light-seed landrace is preferred for consumption due to its superior flavor and less cooking time, while the dark-seeded landrace is used for the treatment of impotence. Curiously, different African tribes use banbara nut for different purposes. For instance, some tribes in Kenya make use of it to treat diarrhoea and nausea [44], in Nigeria it is to treat venereal diseases [40], Pregnant women's nausea and vomiting in South Africa and cataracts in Senegal [40]. In conclusion, banbara nuts serve a variety of purposes, such as supplying minerals and nutrients as functional foods, improving human and animal health (because of their antioxidant qualities), and acting as a commodity to help those in rural sub-Saharan Africa make a living. Therefore, it is necessary to protect and encourage the growth of this crop in order to continue providing the people in the majority of rural African communities with the benefits of it.

#### 4. Nutraceuticals importance of banbara nut

The chairman and founder of the Foundation for Innovation in Medicine, Stephen DeFelice, originated the phrase "nutraceutical" in 1989, combining the words "nutrition" with "pharmaceutical" [45]. DeFelice defines a nutraceutical as a food or component of a food that in addition to offering essential nutrients, has therapeutic benefits, such as the ability to prevent or treat disease or illness [46, 47]. Nutraceuticals can also be defined as non-toxic food ingredients that have been shown by science to have or produce health benefits, like prevention or treatment of disease [48]. The search for potent and innovative antimicrobial chemicals has been spurred by the gradual rise in microorganism resistance to newly developed antibiotics. Researchers have turned their attention to plant-based medicine in their hunt for potent antimicrobial chemicals because of its historical use in the treatment of both infectious and non-infectious disorders, particularly in developing and underdeveloped nations [49]. These medicinal herbs are ingested as teas, elixirs, or juice concoctions for the purpose of treating respiratory infections [50]. Numerous studies [59-53] have demonstrated the importance of these medicinal plants as key sources of chemicals with antibacterial, anticancer, and antioxidant properties. These compounds can be combined to create novel antibiotics that are not resistant to harmful microbes. For instance, Kumar *et al.* [54] investigated the anti-tubercular activity of five plants which are legumes (*Ceasalpinia mimosoides*, *Kingiodendron pinnatum*, *Indigofera cassioides*, *Derris scandens*, and *Humboldtia brunonis*). They found that the leguminous plants contained terpinoids, flavonoids, anthro-quinones, saponins, and phlabotanins. Several studies' results [55, 37, 56] demonstrated that bacteria which are Gram-positive were more vulnerable to plant extracts than Gram-negative bacteria. According to Goli *et al.* [37], the lipopolysaccharides of Gram-negative bacteria's outer membrane are what allow them to withstand plant extracts. But according to a recent study by Ajiboye and Oyejobi [55] employing the method of agar-well diffusion observed that the majority of the extracts were able to stop

Gram-negative bacteria from growing. In his work, Taahir [57] used the direct bioautography method and found that brown and red hulls of banbara nut extracts inhibited Gram-negative bacteria more (*Pseudomonas aeruginosa* ATCC 27853, *pneumoniae* ATCC 700603 and *Klebsiella pneumoniae* subsp.) than bacteria that are Gram-positive bacteria (*Staphylococcus aureus* subsp. *aureus* ATCC 33591), supporting the findings of Ajiboye and Oyejobi [55]. Because brown and red banbara nut hulls linked well with high content of tannin and flavonoid, the ability of the hulls' extracts was attributed to their high content of phenolic compounds [58, 59]. Using the agar diffusion method, Klompong and Benjakul [60] found that extracts from the coat of banbara nuts showed antibacterial activity against wide range of bacteria, which include *Bacillus cereus*, *Escherichia coli*, and *Staphylococcus aureus* as well as *Candida albicans*, *Aspergillus niger*, yeast, and mold, in a manner that is dose-dependent. Using the Kirby-Bauer diffusion method, Anthony [61] discovered that the extracts of banbara nut showed inhibitory activities against *Candida albicans*, *Escherichia coli* *Staphylococcus aureus*, and *Pseudomonas aeruginosa*. All things considered, banbara nuts are a potent antibacterial that can stop prokaryotic and eukaryotic cells from proliferating. The cytoplasmic membrane is a common site of entry for antimicrobial agents, and when this membrane's permeability is disrupted, the membrane itself is destroyed [62]. As a result, the cell deforms, lyses, and dies due to cytoplasmic leakage and coagulation [63]. This substance stops spore germination and mycelial growth in fungus [64]. The phytochemicals found in banbara nuts possess both hydrophilic and hydrophobic properties, which promote their accumulation in cell membranes. This alteration in permeability results in the seepage of intracellular components, weakening of the enzyme systems of microorganisms, and ultimately, their death [63]. It is obvious that the type of organisms being studied and the extract's concentration affect the antibacterial properties of banbara nuts. But no research has determined the exact dosage of banbara nut extracts for treating any human illness in sub-Saharan Africa presently. Banbara nuts have historically been utilized for the treatment of various ailments, and as such, there are numerous chances for additional research on the crop's therapeutic value and potential applications<sup>ss</sup>. Legumes have been found to contain phytochemicals with antioxidant properties by a number of authors [57-73]. Of particular, the hull of the banbara nut has been found to contain phenolic compounds in large quantities, with nutraceutical and functional ingredients than other parts of the plant [68]. It has been observed that the content and kind of phenolics in banbara nuts can be influenced by the pre-processing and extraction methods [72]. Using methanolic extraction optimization, it was found that cooking red banbara nuts improved their nutraceutical profiles by increasing their free radical scavenging speed by a factor of ten when compared to their uncooked counterpart [72]. According to research on the phytochemical components of banbara nuts by Mbagwu *et al.* [74], the crop has more alkaloids than other legumes, and variations in the hulls of different landraces of banbara nuts have been observed [72]. For example, it has been observed that boiling dry beans reduces the phenolic content (protease inhibitors) by 80% to 90%, that treating banbara nut flour with 60% ethanol improves its nutritional quality, and that dehulling can lower the tannin level by up to 92% [80]. Higher concentrations of tanins and flavonoids were discovered in both brown and red

banbara nut landraces in a study intended to determine the therapeutic qualities of banbara nut [72]. The flavonoid compounds found in the hull of brown banbara nuts included higher concentrations of myricetin (1.800 mg g<sup>-1</sup>) and rutin (24.458 mg g<sup>-1</sup>), while the highest tannin compounds found in the hull of red banbara nuts were 0.105 mg g<sup>-1</sup> of ellagic

acid and 0.115 mg g<sup>-1</sup> of chlorogenic acid [58]. Furthermore, two market classes of banbara nuts that are often farmed in Zambia were shown to have polyphenols, particularly higher epicatechin and catechin [72]. Table 1 lists a Some bioactive compounds and chemical components in Bambara nut.

**Table 1:** Some Bioactive compounds and chemical components in Bambara nut.

Type/Class of Nutraceutical	Raw	Cooked	References
<b>Phenolic compounds (mg/g)</b>			
Quercitrin	2.05	1.58	[77]
Rutin	0.427–24.46	3.16	
Quercetin	0.007–6.39	3.94	[77; 78]
Isoquercitrin	0.42	0.29	
Kaempferol	0.052–2.18	3.15	[77; 78]
Myricetin	0.062–1.800	-	
Gallic acid	0.05–1.03	0.41	
Luteolin	1.09	1.67	[77]
Cholorogenic acid	0.03–2.37	0.50	
Catechin	0.01–2.34	-	
Ellagic acid	0.005–1.09	1.42	
Epicatechin	1.15	0.39	
Caffeic acid	-	3.75	[77; 78]
<b>Dietary fiber</b>			
Ursonic acid (% of IDF)	10.6–11.5	-	[79]
Insoluble dietary fibre, IDF (% of TDF)	9.60–10.00	-	
Total dietary fibre, TDF (% of seed)	1.61–10.30	-	[80; 81; 82]
Soluble dietary fiber (% of TDF)	3.00–7.00	-	[82]
Amino acids (% of protein)	3.21–21.38	-	
Glutamic acid	3.21–21.38	-	
Leucine	1.33–10.22	-	[82; 83]
Arginine	1.20–8.25	-	[82; 83]
Lysine	0.99–8.54	-	
Isoleucine	0.89		[83]
<b>Fatty acids (mg/100g)</b>			
Saturated fatty acids	1690	-	
Polyunsaturated fatty (PUFA)	2100	-	
Monounsaturated fatty (MUFA)	1073	-	[83]
<b>Tocopherols and tocotrienol (mg/100g)</b>			
γ-Tocopherol	1.05	-	
α-Tocopherol	0.26	-	[83]
δ-Tocopherol	6.64	-	
α-Tocotrienol	0.10	-	
γ-Tocotrieno	0.18	-	
<b>Phytosterols (% of total sterol)</b>			
β-Sitosterol	1.89–2.23	-	
Campesterol	0.73–3.93	-	[84]
Stigmasterol	0.68–1.78	-	

The authors of that study also demonstrated that heating boosted the nutraceutical and antioxidant properties of these nuts. Additionally, novel phenolic chemicals such as Medioresinol, catechin dimer, GC-hexoside, Quinic acid, catechin glucoside, caffeic acid derivative, p-coumaric acid, and salicylic acid were discovered using HPLC-PDA-ESI-MS profiling in cooked banbara nut seeds [72]. Fascinatingly, when the two market classes of banbara nuts were examined, variations in phenolic compounds were observed [54]. This suggests that there may be room to find more unique phytochemical components of banbara nuts, which might turn the crop into a source of nutraceuticals. The research is lacking on the relationship between the type, content, and medicinal efficacy of banbara nut extracts for the majority of disorders, despite the fact that phenolic chemicals such as flavonoids, alkaloids, lignans, tannins and phenolic acids) are known to treat some illnesses. Moreover, there is a paucity of

research on the impact of extraction methods and pre-processing strategies for optimizing the nutraceutical potential of banbara nuts. Thus, additional research is required to evaluate this crop's potential as an antibacterial and nutraceutical.

### 5. Analysis of Bioactive compounds in banbara nut

The structure, wetness, plasticity, and content of the material will determine the recovery of chemicals from the matrix of the sample. The sample may be fresh or dried, depending on the target compounds. Therefore, it is crucial to prepare the sample matrix before extraction, particularly since certain compounds a ratio of 1:5 to 1:10 solvent/sample (solid vegetal material) for extraction using ultrasonic bath is appropriate [87]. When producing a concentrated extract, such high ratios might be sufficient. To guarantee that, preparation procedures like sifting, drying, and homogenization is

important. By removing interferences, raising the concentration of analyte in the mixture, and producing the ideal size of particle, sample preparation not only maintains the matrix chemicals but also guarantees the extraction efficiency<sup>[85]</sup>. Due to their effect on the cavitation phenomena and the extracts' final concentration, the ratios of sample quantity to solvent and particle size should likewise be put into account in order to optimize extraction yield<sup>[86]</sup>. For the recovery of bioactive chemicals from plants, it has been proposed that the compounds of interest were completely extracted from the matrix of the sample and a larger sample concentration of solvent (1:50, 1:100 and above) would be needed if the extraction's goal is to prepare sample for quantitative analysis of bioactive compounds. The literature reports the solid/solvent ratio ranges; however, it is sometimes unclear if the data are presented based on a dry or wet sample. It is recognized that fresh material, not dry stuff, is typically referred to. To enable the solubilization of the compounds of interest in the case of dry material, the matrix must be hydrated; as a result, the ratio may need to be increased. Sequential extraction procedures can increase efficiency because each extraction will have a new solvent available, improving solubility. Nevertheless, extra procedures like centrifugation or filtration will be needed in between extractions. The major factor in any process of extraction is unquestionably the selection of solvent. The strength and solubility of the matrix interactions with the solvent should be the primary considerations when selecting the extraction solvent. Viscosity, Polarity, specific gravity, surface tension, pH, melting and boiling points, vapor pressure, density, and the impact on the extracted compound's activity and purity are some of the properties of the solvent that need to be monitored<sup>[88]</sup>.

Because they lower the cavitation threshold and make it more difficult to remove the chemicals from the matrix, these aspects should be carefully considered<sup>[89]</sup>. The suitability of the solvent for the extraction parameters, the intermediate and final products to be employed, and the capacity of the solvent to react with the target chemicals under conditions of extraction should all be taken into account. The solvents' physicochemical and biological characteristics are crucial because they not only interact with the extracted substances and treated material but also define the medium's nature. Generally speaking, bioactive chemicals are extracted from plant matrices like herbs, industrial waste, stems, or seeds using organic solvents (methanol, ethanol, acetonitrile, petroleum ether, acetone), water, and combinations of these solvents. Bioactive chemicals from plant sources have long been extracted using organic solvents such ethanol, methanol, acetone, and isopropanol combined with different amounts of water; some extractions employ either 100% organic solvent or water. The stability of the bioactive chemical and the efficacy of the therapies may be significantly impacted by possible modifications to the solvents used during the extraction procedure<sup>[89]</sup>.

According to certain research, acidifying extraction solvents can protect delicate bioactive substances from oxidative deterioration<sup>[90]</sup>. Free radicals that might be created during ultrasonication are stabilized by the hydrogen ions (H<sup>+</sup>) produced by the acids<sup>[89]</sup>. So many researches have shown that organic solvents, such as methanol, very effective at extracting bioactive compounds because of their polarity. In other to reduce the environmental effect of organic solvents while providing comparable or even higher function,

biodegradable and non-toxic substitutes, including ethanol, are being investigated in extraction techniques to some extent<sup>[91]</sup>. New substitutes for harmful solvents include eutectic or ionic solvents that contain the citric acid and lactic acid as well as systems that are multiphasic like cloud point extraction<sup>[92, 95]</sup>. By changing the ionic strength, which impacts the solubility of the chemicals and the sample matrix interactions, the extraction performance can also be affected due to the pH of the solvent. The ideal pH for removing flavonoids from plant matrices has been assessed in a number of research. Mai *et al.*,<sup>[96]</sup> examined how pH of the solvent affected the recovery of flavonoids in *Euonymus alatus* and found that there were high recoveries in acidic pH ranges (2.5–3.5) and fell as the pH ranges increases. The extraction of polyphenols from the peel of pomegranate is influenced by the pH of the solvent, with the optimum outcomes being seen in an acidic medium, according to another study that examined polyphenols in this instance<sup>[97]</sup>. Lower extraction yields were observed at pH values higher than 7.0. Higher flavonoid yields are typically generated in acidic media, according to reports found in the literature. The breakdown of phenolics attached to the polymers carbohydrate and proteins is supported by an acidic pH, which explains this trend for polyphenols<sup>[98]</sup>. At lesser pH values, phenols are protonated which transforms them into molecules that are hydrophobic to interact more strongly with micellar surfactant that are hydrophobic and pass through the micelles easily<sup>[99]</sup>. Higher pH levels allow protons to become more active, deprotonating phenols and making them more ionic. This decreases the solubility of hydrophobic phenolic compounds in micelles. Consequently, more phenols are extracted when the pH is decreased (99; 100).

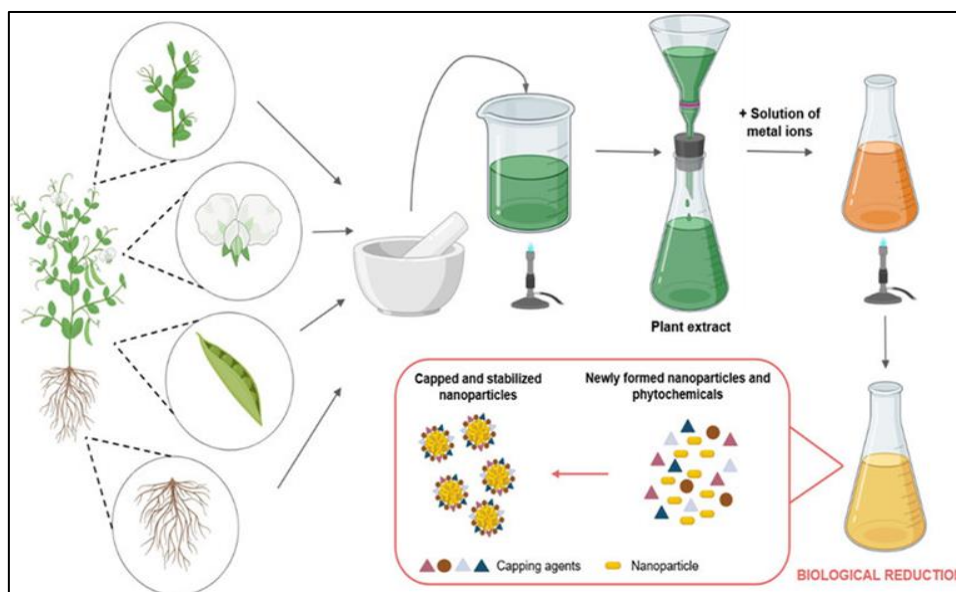
## 6. Plant-mediated nanoparticle synthesis

Numerous bioactive substances present in plants, such as alkaloids, phenols, terpenoids, flavonoids, proteins, steroids, alcohols, saponins function as reducing agents during nanoparticles synthesis. A variety of nanoparticles have been synthesized utilizing plants such as post-harvest leaves of banbara nut, *Ficus benghalensis*, *Acalypha indica*, *Zingiber officinale*, *Centella asiatica*, *Plumbago zeylanica*, *Parthenium hysterophorus*, *Passiflora foetida*, *Sapindus rarak*, *Acalypha indica*, and others<sup>[75, 111, 114]</sup>. Plant extracts are more advantageous than microorganisms when synthesizing green nanoparticles, due to their one-step, nonpathogenic, and economical procedure<sup>[115, 116]</sup>. Bioactive natural compounds have been found to have great potential in this regard and are used to improve the stability, activity, and biodistribution of metal nanoparticles<sup>[118]</sup>, while the extracted bioactive compounds help reduce size and shape-controlled nanoparticles<sup>[119]</sup>. The green chemistry approach has been constantly used for the synthesis of functional nanomaterials to reduce waste, environmental hazards, and the use of toxic chemicals, among other reasons.

In addition to helping the size of the nanoparticles to be fine-tuned, it also helps to remove dangerous byproducts. Numerous studies have demonstrated formulation of nanoparticles employing different parts of plants, such as seeds, callus, fruit, flower, and stem likewise the production using byproducts. The synthesis of nanoparticle using plant extract start by Weighing a precise quantity of plant material in grams, then clean and boil the chosen plant portion in distilled water. Syringe filter tube or a muslin cloth is then used to squeeze and filter the extract. The salt solution that is

added during the filtration process is determined by the type of nanoparticle that will be generated. The solution exhibits a hue shift, signifying the development of nanoparticles that may subsequently be isolated. The main sign indicating that nanoparticles have been synthesized is the noticeable color change (figure 4). If there is no color change in the mixture, it may be because of the medium been high in acidity. To get

around this problem, an alkaline solution can be made by a solution of NaOH. To properly synthesize nanoparticles, salt extract must be applied gradually through a syringe at a moderate volume. The nanoparticle generated may be in the form of colloidal and the synthesis of nanoparticle is verified by a peak at a particular range in nanometer with a decrease in the plasmon absorbance intensity<sup>[117]</sup>.



**Fig 4:** Representation of plant-mediated synthesis of nanoparticles

## 7. Methods of Nanoparticles Synthesis

The most popular technique for nanoparticles synthesis is the Physical (top-down) method, which starts with the fragmentation of bulk material and progresses to the formation of nanoparticles. To create different nanoparticles, the Biological and Chemical (bottom-up) approach collects atoms and molecules. The top-down method greatest advantage is its ability to quickly and efficiently produce a huge number of nanoparticles. Nonetheless, the primary merits of using the bottom-up method are that it produces nanoparticles having distinct crystallographic characteristics and a larger specific surface area<sup>[101]</sup>. It is not possible to synthesize the required shape using a physical method. Nevertheless, unwanted materials can be removed and fewer nanoparticles can be produced by employing a bottom-up strategy. One of the primary techniques for producing nanoparticles utilizing the bottom-up method is the chemical reduction method. Generally speaking, there are three components to the creation of nanoparticles: biological or green, chemical and physical methods<sup>[102]</sup> as shown in figure 4. Furthermore, this technique was proposed as the most important and popular way to create nanoparticles. The physical method uses energy, pressure, and temperature to produce Nanoparticles<sup>[103]</sup>. Atomic condensation, sol-gel, chemical etching, spray-mediated pyrolysis, sputtering and laser pyrolysis are the processes used in the chemical

technique to produce Nanoparticles. Chemical and reaction ratios can change the nanoparticles' morphologies. The produced nanoparticles may face challenges with bioaccumulation, toxicity, regrowth, reuse, and recycling after synthesis<sup>[104,105]</sup>. Green-synthesized Nanoparticles on the other hand, have been shown to be non-toxic<sup>[106]</sup>. Numerous applications in biomaterials research have made use of the green synthesis of nanoparticles with different shapes and sizes<sup>[107,108]</sup>. In the pharmaceutical industry, nanoparticles have been created for a different use, which include the treatment of bacterial and viral illnesses<sup>[109]</sup>. Because it can be produced in large quantities using environmentally friendly methods, the biosynthetic approach offers several advantages over other traditional synthesis protocols. For the amalgamation of nanomaterials, the great bio-diversity and readily available sources of plant output have been extensively studied. These biologically produced nanomaterials have important uses in a number of domains, including treatment, diagnosis, surgical device manipulation, and other product forms. In the treatment of different chronic illnesses, nanomedicine has demonstrated encouraging clinical outcomes. Furthermore, environmentally friendly ways of acquiring Nanoparticles were identified as essential resources for future generations to guard against a number of illnesses<sup>[110]</sup>.

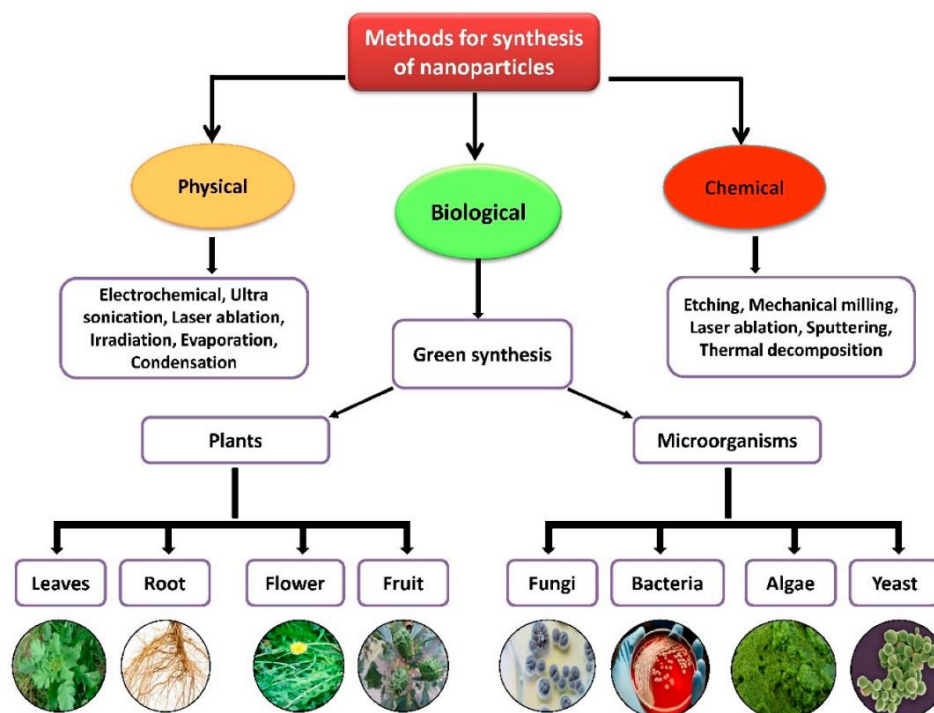


Fig 5: methods for synthesis of Nanoparticle

The use of harmful chemicals in conventional synthesis raises serious concerns since they have a negative effect on plants, soil, and agricultural products when they are discharged into the environment. A volume/high surface ratio is a feature of biological synthesis, and using biological components has

been found to have positive ecological effects. They are regarded as non-toxic since they are known to produce little or no environmental contamination when discharged into the environment (Table 2).

Table 2: Major advantages and disadvantages identified for conventional and biological/green nanomaterials

Materials	Advantages	Disadvantage
Conventional Nanomaterials	<ul style="list-style-type: none"> <li>– NMs are smaller and chemically simpler</li> <li>– They have reliable size, shape, and properties</li> <li>– they follow a standard synthetic process</li> <li>– full-scale use is possible</li> </ul>	<ul style="list-style-type: none"> <li>– The toxicity is unknown when released to the environment</li> <li>– Extremely reactive or potentially unstable</li> <li>– The effect on health of human is unknown</li> <li>– The life cycle of the product is known.</li> </ul>
Biological/green nanomaterials	<ul style="list-style-type: none"> <li>– the procedure for synthesis is eco-friendly</li> <li>– it avoids using solvents that are hazardous</li> <li>– capping agent functions as stabilizers of unstable nanomaterials</li> <li>– it employs Recyclable biological sources</li> </ul>	<ul style="list-style-type: none"> <li>– they are Larger and more chemically complex</li> <li>– feasibility is un-known for large-scale applications</li> <li>– Wide variations in sizes, shapes, and surface properties -</li> <li>– unknown life cycle</li> </ul>

## 5. Uses and application of banbara nut extract in the synthesis of nanoparticles

A study was recently carried out by Ogbuagu *et al.*,<sup>[75]</sup> who green synthesized nanoparticles of silver and zinc oxide utilizing the post-harvest leaves of banbara nut and their antioxidant, antimicrobial and anti-inflammatory potentials. The aqueous leaves extract of the plant was observed to be suitable for the green synthesis of zinc and silver nanoparticles. Due to the reduction of  $\text{Ag}^+$  to  $\text{Ag}^0$ , spherical-shaped leaves nanoparticles with sizes between 20 and 60 nm and rectangular-shaped roots nanoparticles with sizes between 55 and 85 nm were formed. The UV-vis spectroscopy spectroscopic investigation reveals that the nanoparticles' visible section of the spectrum  $\lambda$  peak absorption falls between 434 and 460 nm. The development of amorphous nanoparticles was verified by XRD analysis, which produced diffraction peaks that were not clearly defined. The produced nanoparticles contained functional groups directly involved in the bio-reduction Ag and Zinc

ions to metallic silver and zinc nanoparticles. The produced nanoparticles have extremely low toxicity, according to the toxicity analysis. The results of the antimicrobial activity experiment conducted on *Salmonella typhi* and *Staphylococcus aureus* showed that the produced zinc and silver nanoparticles exhibit antibacterial properties<sup>[75]</sup>. These nanoparticles may be employed in treatments for illnesses brought on by bacteria and other microbes. The nanoparticles may find use in anti-inflammatory therapy as the anti-inflammatory analysis shown high activity for increasing concentrations. At various doses, the activity of anti-oxidant utilizing 2,2-diphenyl-2-picrylhydrazyl (DPPH) demonstrated notable scavenging action. These plants could be used because they are rich in bioactive compounds. Recently research conducted by Eze *et al.*,<sup>[76]</sup> examined the effect of nano-structured bambara nut shell as filler on the mechanical, morphological and physical properties of epoxy matrix. The amount of bambara nut shell fillers in the epoxy matrix has a major impact on the mechanical and physical

properties of the matrix. The ideal composite characteristics were attained at filler loadings up to 15 weight percent, within the parameters of variance in this investigation. Most properties were shown to decline above this filler threshold load. With 15 weight percent of the nano filler, the maximum tensile strength of 80.20 MPa was achieved; however, when the filler loading was increased to 20 weight percent, the strength performance significantly decreased, reaching as low as 39.5 MPa. In a similar vein, the modulus of elasticity decreased for both the 15 and 10 weight percent, going from 138.5 MPa to 118 MPa. It is evident from other mechanical characteristics examined in this study, such as flexural strength and impact energy, that the ideal bambara nut shell nano filler for epoxy matrix is not greater than 15% weight percent. Physical characteristics such as the composite's density showed its highest value at 5 weight percent (0.8g/cm<sup>3</sup>) filler loading. As the filler volume increased above 5 weight percent, composites with lower densities resulted, indicating that the Bambara nut shell nano filler is a light-weight agricultural material appropriate for uses requiring both high strength and low weight. This implies that the composite can be used for both residential and industrial applications at the ideal filler loading, particularly in situations where light weight, high impact energy, and good tensile strength are important considerations<sup>[76]</sup>.

## 6. Other potential applications of bambara nut-derived nanoparticle

Due to their numerous applications in industries, electronics, the environment, energy, and, more specifically, biomedical fields, NPs such as the well-known Ag and Au NPs have been thoroughly studied in this field and are highly intriguing for biological applications. In general, plant-derived green NPs are also less likely to cause serious side effects in humans when compared to chemically synthesized NPs, and they have a wide range of potential uses, including but not limited to:

### Drug Delivery

Therapy Usually entails delivering drugs to a specific target site; if an internal route for drug delivery is not available, external therapeutic methods, such as radiotherapy and surgery, are used. These techniques are frequently used interchangeably or in combination to treat diseases, with the aim of therapy being to always remove the tumors or the cause of illness in a long-lasting manner<sup>[120]</sup>. Nanotechnologies are making a compelling contribution in this area by developing novel drug delivery methods, some of which have been proven effective in a clinical setting and are used in clinical settings<sup>[121]</sup>. For instance, doxorubicin, a drug with high toxicity, can be delivered directly to tumor cells using liposomes (Doxil®) without affecting the kidneys or heart, and paclitaxel incorporated with polymeric mPEG-PLA micelles (GenexolPM®) is used in chemotherapeutic treatment of metastatic breast cancers<sup>[122]</sup>. Improved in vivo distribution, reticuloendothelial system evasion, and favorable pharmacokinetics contribute to the success of nanotechnologies in drug delivery<sup>[123]</sup>. A perfect drug delivery system consists of two components: controlled drug release and targeting ability. By specifically targeting and killing harmful or cancerous cells, side effects can be minimized, and drug efficiency can be ensured. Controlled drug release can also reduce drug side effects<sup>[124]</sup>.

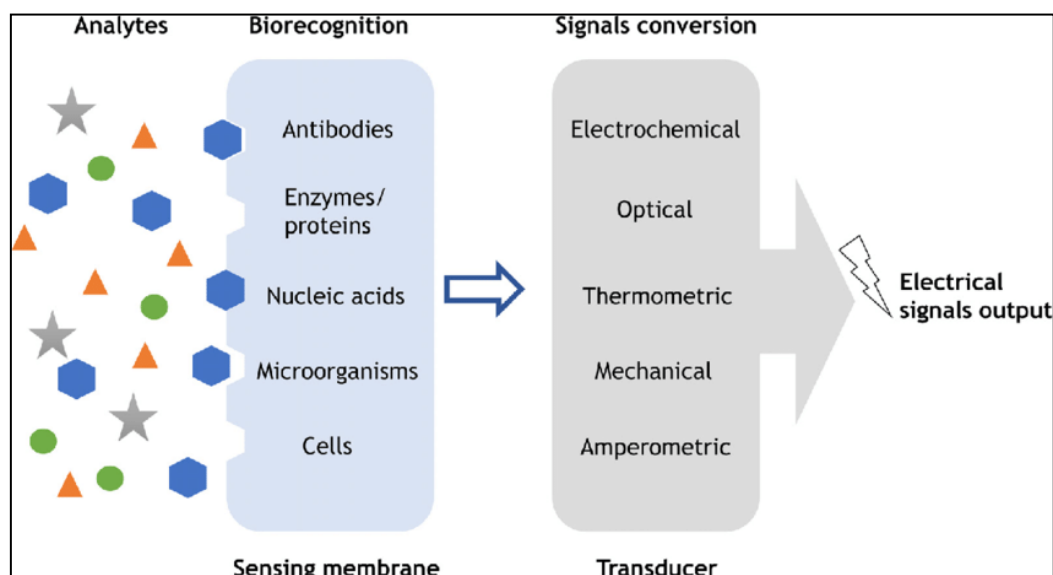
Nanoparticle drug delivery systems have the advantage of reduced irritant reactions and improved penetration within the body due to their small size, allowing for intravenous and other delivery routes. Attaching nano-scaled radioactive anti bodies that complement antigens on the cancer cells with drugs enables the specificity of nanoparticle drug delivery systems. These methods have yielded positive outcomes<sup>[125]</sup>, including enhanced (i) drug bioavailability, (ii) targeted drug delivery, and (iii) uptake of low solubility drugs<sup>[126]</sup>.

### Environment

The environment has been significantly impacted by modern technological innovations. However, advancements in nanotechnology have given us confidence that, with appropriate use, we can restore the ecosystem that was once harmed. By offering rapid and affordable evaluation and management to eliminate contaminants from water, nanoparticles can help address the issue of having clean, accessible water. For instance, a low quantity of copper in drinking water was achieved by using filter paper manufactured from Cu NPs during water purification, which also assisted in reducing the bacteria *E. coli*<sup>[127]</sup>. Cleaning up oil spills has also been shown to benefit from nanoparticles. The carbon particles were able to provide stabilized oil in water emulsions with the aid of tailored surface chemistry<sup>[128]</sup>. Furthermore, in brine shrimp, the carbon black [CB] NPs demonstrated non-toxic effects and the ability to absorb benzene. Since heavy metals like mercury, lead, thallium, and others seriously harm the environment and living things, nanoparticles like carbon nanotubes, nano zeolites, and metal oxides were successful in eliminating them. Nanoscale zero-valent iron is the most practical media since it is inexpensive, readily available, and non-toxic<sup>[129]</sup>. Bimetallic nanoparticles, which combine elemental iron or any other metal with catalysts like silver, gold, nickel, etc., could be used as an alternative to speed up the reduction process.

### Bio-Sensing Applications

Analytical tools called biosensors are used to identify an analyte based on its concentration and occurrence. As seen in figure 6, biosensors are made up of three parts: a transducer, a reader tool, and an element that detects the analyte and produces a signal. A nano-biosensor is a biosensor that functions within the range of nanoscale. They are often used in the clinical setups, for quality assurance, livestock, drug production, biomedical, farming, agricultural, military and defense sectors industrial pollution management, forestry bacterial and virus diagnostics<sup>[149, 150]</sup>. For the benefit of humanity, the utilization of nanoparticles for biological material sensing is highly beneficial<sup>[130]</sup>. These biosensing applications employ a variety of nanoparticles<sup>[131]</sup>. A study demonstrated the application of Green synthesized mesoporous graphene oxide/silica nanocomposites from rich husk ash (RH-GO/SBA-15) for the removal of RhB in aqueous media<sup>[8]</sup>. Another study examined cancer using the green production of Au-Ag alloy mediated by chloroplasts<sup>[132]</sup>. PtNPs mediated by *S. myriocystum* were employed to identify asthma and allergies<sup>[133]</sup>. AuNPs produced by *Hypnea valencia* were utilized to identify pregnant individuals<sup>[134]</sup>. Additionally, the production of AgNPs mediated by *Noctiluca scintillans* was assessed to identify problems with oral discharge and gum disease<sup>[135]</sup>.



**Fig 6:** Schematic illustration of biosensor with its three main components: (a) Sensing membrane (detector), (b) transducer and (c) signal output system.

### Antimicrobial Activity

The scientific community is in danger due to the sharp rise of antibiotic resistance. Thus, it is necessary to create a substance that shows great promise and becomes more effective against strains of bacteria that are resistant to antibiotics. In the past, traditional remedies were made from metals such as iron, copper, silvergold, etc. <sup>[136]</sup>. The researchers used this prior knowledge to determine that nano-based metallic oxide nanoparticles (NPs) have inherent antibacterial activity. Numerous biocidal actions against both Gram-positive and Gram-negative bacteria and eukaryotes were demonstrated by the metallic nanoparticles produced from plants <sup>[137]</sup>. Additionally, it demonstrated that metallic nanoparticles (NPs) effectively inhibited resistant strains, including methicillin-resistant *S. aureus* [MSRA], ampicillin-resistant *E. coli*, and others <sup>[138]</sup>. Currently, plants are used to include metallic nanoparticles' antibacterial properties against bacteria and fungi. Additionally, it is thought that NPs interact with the cell membrane more readily because they have a larger surface-to-volume ratio than their bulk counterparts <sup>[139]</sup>. The cell wall and cell membrane make up the structure of fungi. Phospholipids make up the cell membrane, while mannoproteins,  $\beta$ -1,3-D-glucan and  $\beta$ -1,6-D-glucan proteins, chitin, proteins, lipids, and polysaccharides (such as mannan or galactomannan, glucan, and chitin) make up the cell wall. Through interactions with the cell wall and membrane, the metallic nanoparticles function. Inhibiting  $\beta$ -glucan synthase, a crucial component of the cell wall, comes after the diffusion <sup>[140]</sup>. Oxidative stress, which interacts with macromolecules and results in cell lysis, comes after ROS <sup>[141]</sup>.

### Wound Healing

An injury to the skin tissue caused by trauma or stimulation is called a wound. Numerous treatment approaches are used, including vascular surgery, plant-based therapy, chemotherapeutics, and dressings; however, each method has drawbacks. The use of nanoparticles in wound healing is becoming more popular. It consists of two groups, one of which functions as a delivery agent for repair and the other as a medication that aids in wound healing <sup>[142]</sup>. For example, in albino Wistar male rats' excision wounds. Garg *et al.* <sup>[143]</sup>

showed that silver nanoparticles have antimicrobial and healing properties <sup>[144]</sup>. The silver nanoparticles were synthesized using *Arnebia nobilis* root extract. Rats were given the hydrogel-based formulation. The rats exhibited no harmful effects from NPs, and the cells re-epithelialized. The application of NPs to wound healing is still in its infancy. To fully utilize the therapeutic potential of NPs, more thorough research is necessary to better understand the molecular mechanism and the body's reaction to them.

### Anticancer Property

Because of its high death rate, cancer is one of the primary causes of death. The current chemotherapeutics lose their effectiveness, meaning that the patient develops resistance to the medications after a few cycles. Because of their small size, large surface area, tumor selectivity, and apoptotic activity—that is, their ability to cause cell death and exhibit cytotoxic properties—metallic nanoparticles are showing promise in the treatment of cancer. NPs use a variety of ways to achieve their anticancer effects. The first is apoptosis, in which high ROS levels cause oxidative stress, which in turn causes DNA breakage and cell lysis <sup>[145]</sup>. An additional one is the way that NPs interact with cell membranes and alter cell permeability <sup>[146]</sup>. The silver nanoparticles (PgAg NPs) were synthesized from fresh *Panax ginseng* leaves. In A549, MCF7, and HepG2 cancer cell lines, these nanoparticles showed cytotoxic effects that resulted in oxidative stress <sup>[147]</sup>. A549 cells showed increased phosphorylation of EGF receptors as a result of its inhibition of EGF. Furthermore, PgAg NPs increased the apoptotic process and disrupted the shape of the cells. It is possible that PgAg NPs' anticancer action is due to this interlinkage. The use of NPs to treat cancer is a new field, and further *in vivo* research is needed to fully comprehend the processes that NPs work.

### Vaccines

While inactivated pathogen vaccines usually result in a modest immune response, conventional vaccines based on live-attenuated pathogens carry the danger of reverting to pathogenic virulence. Nanoparticle-based vaccines are a novel strategy that has demonstrated significant promise in overcoming the drawbacks of traditional vaccinations.

Recent developments in chemical and biological engineering have made it possible to precisely manipulate the size, shape, functionality, and surface characteristics of nanoparticles, which improves antigen presentation and boosts immunogenicity.<sup>[148]</sup>

### Other Applications

Nanostructured bambara nut shell as a filler on the mechanical, morphological, and physical characteristics of epoxy matrix is another possible use of this legume plant<sup>[76]</sup>. According to anti-inflammatory study, green synthesized silver and zinc oxide nanoparticles made from post-harvest bambara nut leaves may also be used in anti-inflammatory therapy because their anti-inflammatory properties improve with concentration<sup>[75]</sup>.

### 6. Summary and prospects for research

The benefits of nutraceuticals and the search for greener methods for the synthesis nanoparticles are becoming more widely known, and this has piqued the interest of researchers around the world. Recently, nanoscience has gained researchers' interest due to its significant applications in medical diagnosis, pharmacy, disease curing, electronics, agriculture, space, and chemical industries. The biologically mediated nanoparticles are gotten through plants, bacteria, fungi, actinomycetes, and yeast. Plant extracts are more advantageous than other bio-sources of nanoparticles due to their one-step, non-pathogenic, and economical procedure. Because of bioactive substances found in plants, such as phenols, alkaloids, flavonoids, terpenoids, steroids, saponins, alcohols, and proteins which function as reducing agents and during the synthesis of nanoparticle with less harmful consequences, bio-mediated nanoparticles have been employed extensively to treat a variety of pathogenic disorders. Nanoparticles, which have a number of hazardous qualities for the environment and human health, are produced via non-biological techniques like physical and chemical procedures. Certain characteristics of bio-mediated nanoparticles include increased surface area, increased reactivity, nontoxicity, and biocompatibility. While there hasn't been much research done on the nutraceuticals and synthesis of nanoparticles from bambara nut plants extract, the few researches that has been done has shown the crop's potential as a source of nutraceuticals and nanoparticles due to the presence of bioactive compounds. This review found that the crop has a variety of uses outside of medicine and nutrition. However, very little research has been done on how different nanocomposite are synthesized and their divers' biotechnological applications. Also, there is need to improve the pre-processing techniques, the quantity and quality of phytochemicals including extraction methods for the optimization of the bioactive compounds and the nanoparticles. The majority of the crop's found antimicrobial potentials were mostly against human diseases, indicating that it has the potential to be a natural product that is eventually used to make pharmaceuticals especially when nanoparticles are involved for target delivery and quick response. This review suggest that more investigation is necessary to fully understand the antibacterial potential, nutraceutical benefit and techniques for the synthesis of nanoparticles from bambara nut which is tagged as underutilized legumes with their various applications.

### References

1. Baoua IB, Amadou L, Baributsa D, Murdock LL. Triple bag hermetic technology for post-harvest preservation of Bambara groundnut (*Vigna subterranea* (L.) Verdc.). J Stored Prod Res. 2014;58:48-52. doi:10.1016/j.jspr.2014.02.003
2. Tharanathan RN, Mahadevamma S. Grain legumes—a boon to human nutrition. Trends Food Sci Technol. 2003;14(12):507-18. doi:10.1016/j.tifs.2003.07.002
3. Carbonaro M, Maselli P, Nucara A. Structural aspects of legume proteins and nutraceutical properties. Food Res Int. 2015;76(Pt 1):19-30. doi:10.1016/j.foodres.2014.11.007
4. Pandey M, Verma RK, Saraf SA. Nutraceuticals: new era of medicine. Asian J Pharm Clin Res. 2010;3(3):11-5.
5. Sasi S. Nutraceuticals—a review. Int J Ind Biotechnol Biomat. 2017;3(1):25-9.
6. López-Gutiérrez N, Romero-González R, Plaza-Bolaños P, Vidal JLM, Garrido Frenich A. Identification and quantification of phytochemicals in nutraceutical products from green tea by UHPLC-Orbitrap-MS. Food Chem. 2015;173:607-18. doi:10.1016/j.foodchem.2014.10.092
7. Raveendran P, Fu J, Wallen SL. Completely "green" synthesis and stabilization of metal nanoparticles by using plant extracts. J Am Chem Soc. 2003;125(46):13940-1. doi:10.1021/ja029267j
8. Liou TH, Tseng YK, Liu SM, Lin YT, Wang SY, Liu RT. Green synthesis of mesoporous graphene oxide/silica nanocomposites from rice husk ash: characterization and adsorption performance. Environ Technol Innov. 2021;22:101424. doi:10.1016/j.eti.2021.101424
9. Zeven AC. Landraces: a review of definitions and classifications. Euphytica. 1998;104(2):127-39. doi:10.1023/A:1018683119237
10. Amadou HI, Bebeli PJ, Kaltsikes PJ. Genetic diversity in Bambara groundnut (*Vigna subterranea* L.) germplasm revealed by RAPD markers. Genome. 2001;44(6):995-9. doi:10.1139/g01-096
11. Onimawo IA, Momoh AH, Usman A. Proximate composition and functional properties of four cultivars of Bambara groundnut (*Voandzeia subterranea*). Plant Foods Hum Nutr. 1999;53(2):153-8. doi:10.1023/A:1008027030095
12. Brough SH, Azam-Ali SN, Taylor AJ. The potential of Bambara groundnut (*Vigna subterranea*) in vegetable milk production and basic protein functionality systems. Food Chem. 1993;47(3):277-83. doi:10.1016/0308-8146(93)90161-8
13. Baryeh EA. Physical properties of Bambara groundnuts. J Food Eng. 2001;47(4):321-6. doi:10.1016/S0260-8774(00)00136-9
14. Tibe O, Amarteifio JO, Njogu RM. Trypsin inhibitor activity and condensed tannin content in Bambara groundnut (*Vigna subterranea* (L.) Verdc) grown in southern Africa. J Appl Sci Environ Manag. 2007;11(4):159-64.
15. Mohale KC, Belane AK, Dakora FD. Symbiotic N nutrition, C assimilation, and plant water use efficiency in Bambara groundnut (*Vigna subterranea* L. Verdc)

- grown in farmers' fields in South Africa, measured using  $^{15}\text{N}$  and  $^{13}\text{C}$  natural abundance. *Biol Fertil Soils*. 2014;50(3):307-19. doi:10.1007/s00374-013-0841-3
16. Jideani VA, Mpotokwane SM. Modeling of water absorption of Botswana Bambara varieties using Peleg's equation. *J Food Eng*. 2009;92(2):182-8. doi:10.1016/j.jfoodeng.2008.10.031
  17. Adu-Dapaah HK, Sangwan RS. Improving Bambara groundnut productivity using gamma irradiation and in vitro techniques. *Afr J Biotechnol*. 2004;3(5):260-5.
  18. Mboosso C, Boulay B, Padulosi S, Meldrum G, Mohamadou Y, Niang AB, et al. Fonio and Bambara groundnut value chains in Mali: issues, needs, and opportunities for their sustainable promotion. *Sustainability*. 2020;12(11):4766. doi:10.3390/su12114766
  19. Mubaiwa J, Fogliano V, Chidewe C, Linnemann AR. Bambara groundnut (*Vigna subterranea* (L.) Verdc.) flour: a functional ingredient to favour the use of an unexploited sustainable protein source. *PLoS One*. 2018;13(10):e0205776. doi:10.1371/journal.pone.0205776
  20. Mpotokwane SM, Gaditlhatlhelwe E, Sebaka A, Jideani VA. Physical properties of Bambara groundnuts from Botswana. *J Food Eng*. 2008;89(1):93-8. doi:10.1016/j.jfoodeng.2008.04.017
  21. Eltayeb ARSM, Ali AO, Abou-Arab AA, Abu-Salem FM. Chemical composition and functional properties of flour and protein isolate extracted from Bambara groundnut (*Vigna subterranea*). *Afr J Food Sci*. 2011;5(2):82-90.
  22. Jideani VA, Diedericks CF. Nutritional, therapeutic, and prophylactic properties of *Vigna subterranea* and *Moringa oleifera*. In: Oguntibeju O, ed. *Antioxidant-Antidiabetic Agents and Human Health*. IntechOpen; 2014:187-201. doi:10.5772/57348
  23. Nyau V, Prakash S, Rodrigues J, Farrant J. Identification of nutraceutical phenolic compounds in Bambara groundnuts (*Vigna subterranea* L. Verdc) by HPLC-PDA-ESI-MS. *Br J Appl Sci Technol*. 2015;6(1):77-85. doi:10.9734/BJAST/2015/13611
  24. Okonkwo SI, Opara MF. The analysis of Bambara nut (*Voandzeia subterranea* (L.) thouars) for sustainability in Africa. *Res J Appl Sci*. 2010;6(7):394-6.
  25. Stephens JM. Bambara groundnut—*Voandzeia subterranea* (L.) Thouars. University of Florida IFAS Extension. 2012. EDIS HS547.
  26. Heller J, Begemann F, Mushonga J, eds. Bambara groundnut *Vigna subterranea* (L.) Verdc. Promoting the conservation and use of underutilised and neglected crops. 9. Proceedings of the Workshop on Conservation and Improvement of Bambara Groundnut (*Vigna subterranea* (L.) Verdc.); 1995 Nov 14-16; Harare, Zimbabwe. IPGRI; 1997. p. 14-16.
  27. Basu SN, Roberts JA, Azam-Ali SN, Mayes S. Bambara groundnut. In: Kole C, ed. *Genome Mapping and Molecular Breeding in Plants. Volume 3: Pulses, Sugar and Tuber Crops*. Springer; 2007:157-73. doi:10.1007/978-3-540-34516-9\_10
  28. Massawe FJ, Dickinson M, Roberts JA, Azam-Ali SN. Genetic diversity in Bambara groundnut (*Vigna subterranea* (L.) Verdc) landraces revealed by AFLP markers. *Genome*. 2002;45(6):1175-80. doi:10.1139/g02-093
  29. Mwale SS, Azam-Ali SN, Massawe FJ. Growth and development of Bambara groundnut (*Vigna subterranea*) in response to soil moisture 1. Dry matter and yield. *Eur J Agron*. 2007;26(4):345-53. doi:10.1016/j.eja.2006.09.007
  30. Sesay A. Influence of flooding on Bambara groundnut (*Vigna subterranea* L.) germination: effect of temperature duration and timing. *Afr J Agric Res*. 2009;4(2):100-6.
  31. Ijarotimi OS, Esho TR. Comparison of nutritional composition and antinutrient status of fermented, germinated and roasted Bambara groundnut seeds (*Vigna subterranea*). *Br Food J*. 2009;111(4):376-86. doi:10.1108/00070700910951515
  32. Mahala AGA, Mohammed AA. Nutritive evaluation of Bambara groundnut (*Vigna subterranean*) pods, seeds and hulls as animal feeds. *J Appl Sci Res*. 2010;6(5):383-6.
  33. Aloba AP. Production and organoleptic assessment of akara from Bambara groundnut (*Voandzeia subterranean* (L.) Thouars). *Plant Foods Hum Nutr*. 1999;53(4):313-20. doi:10.1023/A:1008005119326
  34. Adebawale KO, Afolabi TA, Lawal OS. Isolation, chemical modification and physicochemical characterisation of Bambara groundnut (*Voandzeia subterranean*) starch and flour. *Food Chem*. 2002;78(3):305-11. doi:10.1016/S0308-8146(02)00100-0
  35. Lawal OS, Adebawale KO, Adebawale YA. Functional properties of native and chemically modified protein concentrates from bambarra groundnut. *Food Res Int*. 2007;40(8):1003-11. doi:10.1016/j.foodres.2007.05.011
  36. Nti CA. Effects of Bambara groundnut (*Vigna subterranea*) variety and processing on the quality and consumer appeal for its products. *Int J Food Sci Technol*. 2009;44(12):2234-42. doi:10.1111/j.1365-2621.2009.02064.x
  37. Goli AE, Begemann F, Ng NQ. Characterisation and evaluation of IITA's Bambara groundnut (*Vigna subterranean* (L.) Verdc). In: Heller J, Begemann F, Mushonga J, eds. *Proceedings of the Workshop on Conservation and Improvement of Bambara Groundnut (*Vigna subterranean* (L.) Verdc.)*; 1997 Nov 14-16; Harare, Zimbabwe. IPGRI; 1997:101-18.
  38. Uvere PO, Uwaegbute AC, Adedeji EM. Effect of malting on the milling performance and acceptability of Bambara groundnut (*Voandzeia subterranean* Thouars) seeds and products. *Plant Foods Hum Nutr*. 1999;54(1):49-57. doi:10.1023/A:1008177221438
  39. Linnemann AR. Cultivation of Bambara groundnut (*Vigna subterranean* (L.) Verdc.) in the Western Province, Zambia. Report of Field Study of Tropical Crops. Wageningen Agricultural University; 1990.
  40. Murevanhema YY, Jideani VA. Potential of Bambara groundnut (*Vigna subterranea* (L.) Verdc) milk as a probiotic beverage—a review. *Crit Rev Food Sci Nutr*. 2013;53(9):954-67. doi:10.1080/10408398.2011.574803
  41. White PJ, Xing Y. Antioxidants in food: sources of natural antioxidants. In: Pokorny J, Yanishlieva N, Gordon M, eds. *Antioxidants in Food: Practical Applications*. Woodhead Publishing; 2001:22-48.
  42. Adelakun OE, Oyelade OJ, Ade-Omowaye BIO, Adeyemi IA, Van De Venter M. Chemical composition

- and antioxidative properties of Nigeria okra seed (*Abelmoschus esculentus* Moench) flour. *Food Chem Toxicol.* 2009;47(6):1123-6. doi:10.1016/j.fct.2009.01.036
43. Onyilagha JC, Islam S, Ntamatungiro S. Comparative phytochemistry of eleven species of *Vigna* sp. (Fabaceae). *Biochem Syst Ecol.* 2009;37(1):16-9. doi:10.1016/j.bse.2008.09.011
  44. Mkandawire CH. Review of Bambara groundnut (*Vigna subterranea* (L.) Verdc.) production in sub-sahara Africa. *Agric J.* 2007;2(4):464-70.
  45. Brower V. Nutraceutical poised for a healthy slice of the healthcare market? *Nat Biotechnol.* 1998;16(8):728-31. doi:10.1038/nbt0898-728
  46. DeFelice SL. What is a true nutraceutical? And what is the nature and size of the U.S. Market?. *FIM*; 1994.
  47. Wildman REC, Kelley M. Nutraceuticals and functional foods. In: Wildman REC, ed. *Handbook of Nutraceuticals and Functional Foods*. 2nd ed. CRC Press; 2007:1-22.
  48. Kathleen C, Stephen D. Nutraceuticals: what are they and do they work? Kentucky Enquirer Research Inc; 2009:30-5.
  49. Yagi S, Drouart N, Bourgaud F, Henry M, Chapleur Y, Laurain-Mattar D. Antioxidant and antiglycation properties of Hydnora johannis roots. *S Afr J Bot.* 2013;84:124-7. doi:10.1016/j.sajb.2012.10.003
  50. Rojas JJ, Ochoa VJ, Ocampo SA, Muñoz JF. Screening for antimicrobial activity of ten medicinal plants used in Colombian folkloric medicine: a possible alternative in the treatment of non-nosocomial infections. *BMC Complement Altern Med.* 2006;6:2. doi:10.1186/1472-6882-6-2
  51. Abu-Shanab B, Adwan G, Abu-Safiya D, Jarrar N, Adwan K. Antibacterial activities of some plant extracts utilized in popular medicine in Palestine. *Turk J Biol.* 2004;28(2):99-102.
  52. Patra JK, Dhal NK, Thatoi HN. In vitro bioactivity and phytochemical screening of Suaeda maritima (Dumort): a mangrove associate from Bhitarkanika, India. *Asian Pac J Trop Med.* 2011;4(9):727-34. doi:10.1016/S1995-7645(11)60182-X
  53. Iwu MM, Duncan AR, Okunji CO. New antimicrobials of plant origin. In: Janick J, ed. *Perspectives on New Crops and New Uses*. ASHS Press; 1999:457-62.
  54. Kumar JK, Prasad AGD, Chaturvedi V. Phytochemical screening of five medicinal legumes and their evaluation for in vitro anti-tubercular activity. *Ayu.* 2014;35(1):98-102. doi:10.4103/0974-8520.141952
  55. Ajiboye AA, Oyejobi GK. In vitro antimicrobial activities of *Vigna subterranean*. *J Antimicrob Agents.* 2017;3(1):1-4.
  56. Brantner A, Males Z, Pepeljnjak S, Antolic A. Antimicrobial activity of Paliurus spina-christi Mill. *J Ethnopharmacol.* 1996;52(2):119-22. doi:10.1016/0378-8741(96)01408-0
  57. Taahir H. Bambara groundnut (*Vigna subterranean*) from Mpumalanga Province of South Africa: phytochemical and antimicrobial properties of seeds and product extracts [MSc thesis]. Cape Peninsula University of Technology; 2017.
  58. Celis C, Garcia A, Sequeda G, Mendez G, Torrenegra R. Antimicrobial activity of extracts obtained from Anacardium excelsum against some pathogenic microorganisms. *Emir J Food Agric.* 2011;23(3):249-57.
  59. Carraturo A, Raieta K, Tedesco I, Kim J, Russo GL. Antibacterial activity of phenolic compounds derived from Ginkgo biloba sarcotestas against food-borne pathogens. *Br Microbiol Res J.* 2014;4(1):18-27.
  60. Klompong V, Benjakul S. Antioxidative and antimicrobial activities of the extracts from the seed coat of Bambara groundnut (*Voandzeia subterranea*). *RSC Adv.* 2015;5(15):9973-85. doi:10.1039/C4RA10955D
  61. Anthony WW. Evaluation of phytoconstituents, antioxidants potential, cytotoxic, antimicrobial activities and mineral composition of *Vigna subterranea* (L.) Verdc. extracts [MSc thesis]. Jomo Kenyatta University of Agriculture and Technology; 2018.
  62. Weerakkody NS, Caffin N, Turner MS, Dykes GA. In vitro antimicrobial activity of less-utilized spice and herb extracts against selected food-borne bacteria. *Food Control.* 2010;21(10):1408-14. doi:10.1016/j.foodcont.2010.04.014
  63. Liang FLH, Yuan Q, Li C. In vitro antimicrobial effects and mechanism of action of selected plant essential oil combinations against four food-related microorganisms. *Int J Food Microbiol.* 2011;44(10):3057-64. doi:10.1016/j.ijfoodmicro.2011.07.030
  64. Kannan VR, Sumathi CS, Balasubramanian V, Ramesh N. Elementary chemical profiling and antifungal properties of cashew (*Anacardium occidentale* L.) nuts. *Bot Res Int.* 2009;2(4):253-7.
  65. Cushnie TP, Lamb AJ. Antimicrobial activity of flavonoids. *Int J Antimicrob Agents.* 2005;26(5):343-56. doi:10.1016/j.ijantimicag.2005.09.002
  66. Mølgaard P, Holler JG, Asar B, Liberna I, Rosenbæk LB, Jebjerg CP, Jørgensen L, Lauritzen J, Guzman A, Adersen A, Simonsen HT. Antimicrobial evaluation of Huilliche plant medicine used to treat wounds. *J Ethnopharmacol.* 2011;138(1):219-27. doi:10.1016/j.jep.2011.09.006
  67. Harris T, Jideani V, Le Roes-Hill M. Flavonoids and tannin composition of Bambara groundnut (*Vigna subterranea*) of Mpumalanga, South Africa. *Heliyon.* 2018;4(9):e00833. doi:10.1016/j.heliyon.2018.e00833
  68. Spassieva S, Hille J. Plant sphingolipids today—are they still enigmatic? *Plant Biol.* 2003;5(2):125-36. doi:10.1055/s-2003-39001
  69. Tsamo AT, Ndibewu PP, Dakora FD. Phytochemical profile of seeds from 21 Bambara groundnut landraces via UPLC-qTOF-MS. *Food Res Int.* 2018;112:160-8. doi:10.1016/j.foodres.2018.06.028
  70. Farag MA, Gad HA, Heiss AG, Wessjohann LA. Metabolomics driven analysis of six Nigella species seeds via UPLC-qTOF-MS and GC-MS coupled to chemometrics. *Food Chem.* 2014;151:333-42. doi:10.1016/j.foodchem.2013.11.032
  71. Nyau V, Prakash S, Rodrigues J, Farrant J. Domestic cooking effects of Bambara groundnuts and common beans in the antioxidant properties and polyphenol profiles. *J Food Res.* 2017;6(2):24-37. doi:10.5539/jfr.v6n2p24
  72. Kumar N, Pruthi V. Potential applications of ferulic acid from natural sources. *Biotechnol Rep.* 2014;4:86-93. doi:10.1016/j.btre.2014.09.002
  73. Mbagwu FN, Okafor VU, Ekeanyanwu J. Phytochemical screening on four edible legumes (*Vigna subterranean*, *Glycine max*, *Arachis hypogea*, and *Vigna unguiculata*)

- found in eastern Nigeria. *Afr J Plant Sci.* 2011;5(7):370-2.
74. Ogbuagu AS, Innocent LC, Okoye NN, Umeh SO, Ogbuagu JO. Green synthesis of silver and zinc oxide nanoparticles using post-harvest leaves of *Vigna subterranean* and their antimicrobial, anti-inflammatory and antioxidant potentials. *Asian J Appl Chem Res.* 2023;13(3):1-14. doi:10.9734/ajacr/2023/v13i3243
  75. Eze WU, Yakubu MK, Buba MA, Kuzmin A, Santos-Ndukwe IB, Ugbaja MI, Bayero AH. Effect of nano-structured Bambara nut shell (*Vigna subterranea* (L.) Verdc) as filler on the physical, mechanical and morphological properties of epoxy matrix. *J Mater Environ Sci.* 2022;13(10):1155-70.
  76. Salawu S. Comparative study of the antioxidant activities of methanolic extract and simulated gastrointestinal enzyme digest of Bambara nut (*Vigna subterranean*). *FUTA J Res Sci.* 2016;1(1):107-20.
  77. Harris T, Jideani V, Le Roes-Hill M. Flavonoids and tannin composition of Bambara groundnut (*Vigna subterranea*) of Mpumalanga, South Africa. *Heliyon.* 2018;4(9):e00833. doi:10.1016/j.heliyon.2018.e00833
  78. Maphosa Y, Jideani VA. Physicochemical characteristics of Bambara groundnut dietary fibres extracted using wet milling. *S Afr J Sci.* 2016;112(1/2):1-8. doi:10.17159/sajs.2016/20150094
  79. Mazahib AM, Nuha MO, Salawa IS, Babiker EE. Some nutritional attributes of Bambara groundnut as influenced by domestic processing. *Int Food Res J.* 2013;20(3):1165-71.
  80. Abiodun AO, Adepeju AB. Effect of processing on the chemical, pasting and antinutritional composition of Bambara nut (*Vigna Subterranea* L. Verdc) flour. *Adv J Food Sci Technol.* 2011;3(4):224-7.
  81. Yao DN, Kouassi KN, Erba D, Scazzina F, Pellegrini N, Casiraghi MC. Nutritive evaluation of the Bambara groundnut Ci12 landrace [*Vigna subterranea* (L.) verdc. (Fabaceae)]. *Int J Mol Sci.* 2015;16(9):21428-41. doi:10.3390/ijms160921428
  82. Baptista A, Pinho O, Pinto E, Casal S, Mota C, Ferreira IMPLVO. Characterization of protein and fat composition of seeds from common beans (*Phaseolus vulgaris* L.), cowpea (*Vigna unguiculata* L. Walp) and Bambara groundnuts (*Vigna subterranea* L. Verdc) from Mozambique. *J Food Meas Charact.* 2017;11(2):442-50. doi:10.1007/s11694-016-9412-2
  83. Adeyeye EI, Olaleye AA, Adesina AJ. Lipid composition of testa, dehulled seed and whole seed of Bambara groundnut (*Vigna subterranea* L. Verdc). *Curr Adv Plant Sci Res.* 2015;2(1):1-9.
  84. Rostagno MA, Villares A, Guillamón E, García-Lafuente A, Martínez JA. Sample preparation for the analysis of isoflavones from soybeans and soy foods. *J Chromatogr A.* 2009;1216(1):2-29. doi:10.1016/j.chroma.2008.11.035
  85. Vilkhuk K, Manasseh R, Mawson R, Ashokkumar M. Ultrasonic recovery and modification of food ingredients. In: Feng H, Barbosa-Cánovas GV, Weiss J, eds. *Ultrasound Technologies for Food and Bioprocessing.* Springer; 2011:345-68. doi:10.1007/978-1-4419-7472-3\_13
  86. Vinatoru M, Mason TJ, Calinescu I. Ultrasonically assisted extraction (UAE) and microwave assisted extraction (MAE) of functional compounds from plant materials. *TrAC Trends Anal Chem.* 2017;97:159-78. doi:10.1016/j.trac.2017.09.002
  87. Mason TJ, Lorimer JP. *Applied Sonochemistry: The Uses of Power Ultrasound in Chemistry and Processing.* Wiley-VCH; 2002. doi:10.1002/352760054X
  88. Dzah CS, Duan Y, Zhang H, Wen C, Zhang J, Chen G, Ma H. The effects of ultrasound assisted extraction on yield, antioxidant, anticancer and antimicrobial activity of polyphenol extracts: a review. *Food Biosci.* 2020;35:100547. doi:10.1016/j.fbio.2020.100547
  89. Dzah CS. Influence of fruit maturity on antioxidant potential and chilling injury resistance of peach fruit (*Prunus persica*) during cold storage. *Afr J Food Agric Nutr Dev.* 2014;14(6):9578-91.
  90. Fu X, Belwal T, Cravotto G, Luo Z. Sono-physical and sono-chemical effects of ultrasound: primary applications in extraction and freezing operations and influence on food components. *Ultrason Sonochem.* 2020;60:104726. doi:10.1016/j.ultsonch.2019.104726
  91. Vankar PS, Srivastava J. Ultrasound-assisted extraction in different solvents for phytochemical study of *Canna indica*. *Int J Food Eng.* 2010;6(3). doi:10.2202/1556-3758.1599
  92. Ekezie FGC, Sun DW, Cheng JH. Acceleration of microwave assisted extraction processes of food components by integrating technologies and applying emerging solvents: a review of latest developments. *Trends Food Sci Technol.* 2017;67:160-72. doi:10.1016/j.tifs.2017.06.006
  93. Cunha SC, Fernandes JO. Extraction techniques with deep eutectic solvents. *TrAC Trends Anal Chem.* 2018;105:225-39. doi:10.1016/j.trac.2018.05.001
  94. Biata NR, Mashile GP, Ramontja J, Mketo N, Nomngongo PN. Application of ultrasound-assisted cloud point extraction for preconcentration of antimony, tin and thallium in food and water samples prior to ICP-OES determination. *J Food Compos Anal.* 2019;76:14-21. doi:10.1016/j.jfca.2018.11.004
  95. Mai X, Liu Y, Wang TX, Wang L, Lin Y, Zeng H, et al. Sequential extraction and enrichment of flavonoids from *Euonymus alatus* by ultrasonic-assisted polyethylene glycol-based extraction coupled to temperature-induced cloud point extraction. *Ultrason Sonochem.* 2020;66:105073. doi:10.1016/j.ultsonch.2020.105073
  96. Motikar PD, More PR, Arya SS. A novel, green environment friendly cloud point extraction of polyphenols from pomegranate peels: a comparative assessment with ultrasound and microwave-assisted extraction. *Sep Sci Technol.* 2020;56(6):1014-25. doi:10.1080/01496395.2020.1746969
  97. Ilbay Z, Sahin S, Büyükkabasakal K. A novel approach for olive leaf extraction through ultrasound technology: response surface methodology versus artificial neural networks. *Korean J Chem Eng.* 2014;31(9):1661-7. doi:10.1007/s11814-014-0106-3
  98. El-Abbassi A, Kiai H, Raiti J, Hafidi A. Cloud point extraction of phenolic compounds from pretreated olive mill wastewater. *J Environ Chem Eng.* 2014;2(3):1480-6. doi:10.1016/j.jece.2014.06.024
  99. Gortzi O, Lalas S, Chatzilazarou A, Katsoyannos E, Papaconstantinou S, Dourtoglou E. Recovery of natural antioxidants from olive mill waste water using Genapol-X080. *J Am Oil Chem Soc.* 2008;85(2):133-40. doi:10.1007/s11746-007-1180-z

100. Krukowski S, Lysenko N, Kolodziejski W. Synthesis and characterization of nanocrystalline composites containing calcium hydroxyapatite and glycine. *J Solid State Chem.* 2018;264:59-67. doi:10.1016/j.jssc.2018.05.008
101. Ramanathan S, Gopinath SCB, Arshad MKM, Poopalan P, Perumal V. Nanoparticle synthetic methods: strength and limitations. In: Gopinath SCB, Gang F, eds. *Nanoparticles in Analytical and Medical Devices*. Elsevier; 2021:31-43. doi:10.1016/B978-0-12-821163-2.00005-7
102. Mittal AK, Chisti Y, Banerjee UC. Synthesis of metallic nanoparticles using plant extracts. *Biotechnol Adv.* 2013;31(2):346-56. doi:10.1016/j.biotechadv.2013.01.003
103. Marchiol L. Synthesis of metal nanoparticles in living plants. *Ital J Agron.* 2012;7(4):e27. doi:10.4081/ija.2012.e27
104. Qi P, Wang N, Zhang T, Feng Y, Zhou X, Zeng D, et al. Anti-virulence strategy of novel dehydroabietic acid derivatives: design, synthesis, and antibacterial evaluation. *Int J Mol Sci.* 2023;24(3):2897. doi:10.3390/ijms24032897
105. Sobczak-Kupiec A, Pluta K, Drabczyk A, Wlo's M, Tyliszczak B. Synthesis and characterization of ceramic-polymer composites containing bioactive synthetic hydroxyapatite for biomedical applications. *Ceram Int.* 2018;44(11):13630-8. doi:10.1016/j.ceramint.2018.04.181
106. Gong D, Sun L, Li X, Zhang W, Zhang D, Cai J. Micro/nanofabrication, assembly, and actuation based on microorganisms: recent advances and perspectives. *Small Struct.* 2023;4(3):2200356. doi:10.1002/sstr.202200356
107. Gong D, Celi N, Zhang D, Cai J. Magnetic biohybrid microrobot multimers based on *Chlorella* cells for enhanced targeted drug delivery. *ACS Appl Mater Interfaces.* 2022;14(5):6320-30. doi:10.1021/acsami.1c19663
108. EmamiMoghaddam SA, Ghadam P, Rahimzadeh F. Biosynthesis of cadmium sulfide nanoparticles using aqueous extract of *Lactobacillus acidophilus* along with its improvement by response surface methodology. *J Clean Prod.* 2022;356:131848. doi:10.1016/j.jclepro.2022.131848
109. Elizabeth MK, Devi RU, Raja KP, Krishna KB. Synthesis of phyto based metal nanoparticles: a green approach. *J Pharm Res Int.* 2022;34(1):20-32. doi:10.9734/jpri/2022/v34i130605
110. Verma Y, Singh SK, Jatav HS, Rajput VD, Minkina T. Interaction of zinc oxide nanoparticles with soil: insights into the chemical and biological properties. *Environ Geochem Health.* 2021;43(12):1-14. doi:10.1007/s10653-021-01020-6
111. Ikram M, Javed B, Raja NI, Mashwani ZUR. Biomedical potential of plant-based selenium nanoparticles: a comprehensive review on therapeutic and mechanistic aspects. *Int J Nanomed.* 2021;16:249. doi:10.2147/IJN.S288021
112. Peralta-Videa JR, Huang Y, Parsons JG, Zhao L, Lopez-Moreno L, Hernandez-Viezcas JA, Gardea-Torresdey JL. Plant-based green synthesis of metallic nanoparticles: scientific curiosity or a realistic alternative to chemical and physical methods? *Nanotechnol Environ Eng.* 2016;1(1):1-29. doi:10.1007/s41204-016-0005-4
113. Wang C, Kim YJ, Singh P, Mathiyalagan R, Jin Y, Yang DC. Green synthesis of silver nanoparticles by *Bacillus methylotrophicus*, and their antimicrobial activity. *Artif Cells Nanomed Biotechnol.* 2016;44(4):1127-32. doi:10.3109/21691401.2015.1013955
114. Roy A. Plant derived silver nanoparticles and their therapeutic applications. *Curr Pharm Biotechnol.* 2021;22(14):1834-47. doi:10.2174/1389201022666211027155708
115. Nagore P, Ghotekar S, Mane K, et al. Structural properties and antimicrobial activities of *Polyalthia longifolia* leaf extract-mediated CuO nanoparticles. *BioNanoScience.* 2021;11:579-89. doi:10.1007/s12668-021-00851-4
116. Kaur S, Setia G, Sikenis M, Kumar S. Synthesis of biogenic nanomaterials, their characterization, and applications. In: Kaur S, ed. *Biogenic Nano-Particles and Their Use in Melio-Rational Design of Medical Devices*. Springer; 2024:45-75.
117. Akinfenwa AO, Abdul NS, Docrat FT, Marnewick JL, Luckay RC, Hussein AA. Cytotoxic effects of phytomediated silver and gold nanoparticles synthesised from *Rooibos* (*Aspalathus linearis*), and *Aspalathin*. *Plants.* 2021;10(11):2460. doi:10.3390/plants10112460
118. Saravanan M, Barabadi H, Vahidi H. Green nanotechnology: isolation of bioactive molecules and modified approach of biosynthesis. In: Gopinath SCB, ed. *Biogenic Nanoparticles for Cancer Theranostics*. Elsevier; 2021:101-22. doi:10.1016/B978-0-12-821467-1.00005-7
119. Liang J-L, Luo G-F, Chen W-H, Zhang X-Z. Recent advances in engineered materials for immunotherapy-involved combination cancer therapy. *Adv Mater.* 2021;33(31):2007630. doi:10.1002/adma.202007630
120. Saeedi M, Eslamifar M, Khezri K, Dizaj SM. Applications of nanotechnology in drug delivery to the central nervous system. *Biomed Pharmacother.* 2019;111:666-75. doi:10.1016/j.biopha.2018.12.133
121. Lombardo D, Kiselev MA, Caccamo MT. Smart nanoparticles for drug delivery application: development of versatile nanocarrier platforms in biotechnology and nanomedicine. *J Nanomater.* 2019;2019:3702518.
122. Ferreira CA, Goel S, Ehlerding EB, Rosenkrans ZT, Jiang D, Sun T, et al. Ultrasmall porous silica nanoparticles with enhanced pharmacokinetics for cancer theranostics. *Nano Lett.* 2021;21(11):4692-9. doi:10.1021/acs.nanolett.1c00895
123. Singh AP, Biswas A, Shukla A, Maiti P. Targeted therapy in chronic diseases using nanomaterial-based drug delivery vehicles. *Signal Transduct Target Ther.* 2019;4(1):33. doi:10.1038/s41392-019-0068-3
124. Suri SS, Fenniri H, Singh B. Nanotechnology-based drug delivery systems. *J Occup Med Toxicol.* 2007;2(1):16. doi:10.1186/1745-6673-2-16
125. Mazayen ZM, Ghoneim AM, Elbatany RS, Basalious EB, Bendas ER. Pharmaceutical nanotechnology: from the bench to the market. *Future J Pharm Sci.* 2022;8(1):12. doi:10.1186/s43094-022-00400-0
126. Rodd AL, Creighton MA, Vaslet CA, Rangel-Mendez JR, Hurt RH, Kane AB. Effects of surface-engineered nanoparticle-based dispersants for marine oil spills on the model organism *Artemia franciscana*. *Environ Sci*

- Technol. 2014;48(11):6419-27. doi:10.1021/es500892m
127. Fu F, Dionysiou DD, Liu H. The use of zero-valent iron for groundwater remediation and wastewater treatment: a review. *J Hazard Mater.* 2014;267:194-205. doi:10.1016/j.jhazmat.2013.12.062
  128. Galib, Barve M, Mashru M, Jagtap C, Patgiri BJ, Prajapati PK. Therapeutic potentials of metals in ancient India: a review through Charaka Samhita. *J Ayurveda Integr Med.* 2011;2(2):55-63. doi:10.4103/0975-9476.82523
  129. Michael H, Alan LG, Serge C. Nanomaterials for biosensing applications: a review. *Front Chem.* 2014;2:63. doi:10.3389/fchem.2014.00063
  130. Ramesh M, Janani R, Deepa C, Rajeshkumar L. Nanotechnology-enabled biosensors: a review of fundamentals, design principles, materials, and applications. *Biosensors.* 2023;13(1):40. doi:10.3390/bios13010040
  131. Zhang Y, Gao G, Qian Q, Cui D. Chloroplasts-mediated biosynthesis of nanoscale Au-Ag alloy for 2-butanone assay based on electrochemical sensor. *Nanoscale Res Lett.* 2012;7(1):475. doi:10.1186/1556-276X-7-475
  132. Sharma D, Kanchi S, Bisetty K. Biogenic synthesis of nanoparticles: a review. *Arab J Chem.* 2019;12(8):3576-600. doi:10.1016/j.arabjc.2014.12.008
  133. Kuppusamy P, Mashitah MY, Maniam GP, Govindan N. Biosynthesized gold nanoparticle developed as a tool for detection of HCG hormone in pregnant women urine sample. *Asian Pac J Trop Dis.* 2014;4(3):237-41. doi:10.1016/S2222-1808(14)60570-8
  134. Elgamouz A, Idriss H, Nassab C, Bihi A, Bajou K, Hasan K, Abu Haija M, Patole SP. Green synthesis, characterization, antimicrobial, anti-cancer, and optimization of colorimetric sensing of hydrogen peroxide of algae extract capped silver nanoparticles. *Nanomaterials.* 2020;10(10):1861. doi:10.3390/nano10101861
  135. Eltarahony M, Zaki S, Elkady M, Abd-El-Haleem D. Biosynthesis, characterization of some combined nanoparticles, and its biocide potency against a broad spectrum of pathogens. *J Nanomater.* 2018;2018:5263814. doi:10.1155/2018/5263814
  136. Baptista PV, McCusker MP, Carvalho A, Ferreira DA, Mohan NM, Martins M, Fernandes AR. Nano-strategies to fight multidrug resistant bacteria—"a battle of the titans". *Front Microbiol.* 2018;9:1441. doi:10.3389/fmicb.2018.01441
  137. Slavin YN, Asnis J, Häfeli UO, Bach H. Metal nanoparticles: understanding the mechanisms behind antibacterial activity. *J Nanobiotechnol.* 2017;15(1):1-20. doi:10.1186/s12951-017-0308-z
  138. Xia T, Kovochich M, Liong M, Mädler L, Gilbert B, Shi H, Yeh JI, Zink JI, Nel AE. Comparison of the mechanism of toxicity of zinc oxide and cerium oxide nanoparticles based on dissolution and oxidative stress properties. *ACS Nano.* 2008;2(10):2121-34. doi:10.1021/nn800511k
  139. Arciniegas-Grijalba PA, Patiño-Portela MC, Mosquera-Sánchez LP, Guerrero-Vargas JA, Rodríguez-Páez JE. ZnO nanoparticles (ZnO-NPs) and their antifungal activity against coffee fungus *Erythricium salmonicolor*. *Appl Nanosci.* 2017;7(5):225-41. doi:10.1007/s13204-017-0561-3
  140. Zeng R, Dong J. The Hippo signaling pathway in drug resistance in cancer. *Cancers.* 2021;13(2):318. doi:10.3390/cancers13020318
  141. Hamdan S, Pastar I, Drakulich S, Dikici E, Tomic-Canic M, Deo S, Daunert S. Nanotechnology-driven therapeutic interventions in wound healing: potential uses and applications. *ACS Cent Sci.* 2017;3(3):163-75. doi:10.1021/acscentsci.6b00371
  142. Garg S, Chandra A, Mazumder A, Mazumder R. Green synthesis of silver nanoparticles using *Arnebia nobilis* root extract and wound healing potential of its hydrogel. *Asian J Pharm.* 2014;9(1):42-7. doi:10.4103/0973-8398.134925
  143. Naraginti S, Kumari PL, Das RK, Sivakumar A, Patil SH, Andhalkar VV. Amelioration of excision wounds by topical application of green synthesized, formulated silver and gold nanoparticles in albino Wistar rats. *Mater Sci Eng C Mater Biol Appl.* 2016;62:293-300. doi:10.1016/j.msec.2016.01.069
  144. Lim ZZJ, Li J, Ng CT, Yung LYL, Bay BH. Gold nanoparticles in cancer therapy. *Acta Pharmacol Sin.* 2011;32(8):983-90. doi:10.1038/aps.2011.82
  145. Castro-Aceituno V, Ahn S, Simu SY, Singh P, Mathiyalagan R, Lee HA, Yang DC. Anticancer activity of silver nanoparticles from *Panax ginseng* fresh leaves in human cancer cells. *Biomed Pharmacother.* 2016;84:158-64. doi:10.1016/j.biopha.2016.09.016
  146. Nethi SK, Das S, Patra CR, Mukherjee S. Recent advances in inorganic nanomaterials for wound-healing applications. *Biomater Sci.* 2019;7(7):2652-74. doi:10.1039/C9BM00423H
  147. Al-Halifa S, Gauthier L, Arpin D, Bourgault S, Archambault D. Nanoparticle-based vaccines against respiratory viruses. *Front Immunol.* 2019;10:22. doi:10.3389/fimmu.2019.00022
  148. Seo Y, Jeong J, Lee J, Choi HS, Kim J, Lee H. Innovations in biomedical nanoengineering: nanowell array biosensor. *Nano Convergence.* 2018;5(1):7. doi:10.1186/s40580-018-0141-6
  149. Singh P, Pandey SK, Singh J, Srivastava S, Sachan S, Singh SK. Biomedical perspective of electrochemical nanobiosensor. *Nano-Micro Lett.* 2016;8(3):193-203. doi:10.1007/s40820-015-0077-x
  150. Amusan O, Ogunbiyi OJ, Shoge MO, Jemkur M, Joseph PS. Evaluation of phytochemical compounds and proximate analysis of doum palm fruit (*Hyphaene thebaica*) blend with turmeric powder (*Curcuma longa*). *BMC Chem.* 2024;18(1):140. doi:10.1186/s13065-024-01256-6